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FBI pH 5.5	Clone	Α̈́	Vmax	Vmax/Km	Vmax/Km X improved	AP1 pH 5.5	Clone	Æ	Vmax	Vmax/Km	X improved	
50-225 uM [S]	4F13 G12	62	15	0.242	7.1	50-225 uM [S]	4F13 G12	365	44.2	0.121	6.1	
	4F15 A11	28	13.6	0.234	6.9		4F15 A11	438	46	0.105	5.3	
	4F15 C3	39	თ	0.231	6.8		4F15 C3	563	27.6	0.049	2.5	
	4F6 A11 4F3 B5	191 101.8	28.3 16.4	0.148	4.4		4F6 A11 4F3 B5	288 378	28.3 25.3	0.098	9.4 9.5	
·	4F2 G10 4F19 F2 4F21 C8 4F22 B2	41.2 235 113 161	9.4 59.5 22 21.5	0.228 0.253 0.195 0.134	6.7 7.4 5.7 3.9		4F2 G10 4 4F19 F2 4 4F21 C8 4F22 B2	2209 652 305.5 444	61.3 18 55.9 64.5	0.028 0.028 0.183 0.145	1.4 1.4 9.2 7.3	
	4F28 G1	172	23.9	0.139	1.1		4F28 G1	9	8	N	ND	
W' 22 hits (>3X Improved)	WT nproved)	349	11.8	0.034	<del></del>	W 11 hits (>3X Improved)	WT nproved)	450	6.2	0.02	<del>-</del>	
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<u>Н</u>

Results: Kinetic parameters of pH-optimized candidates

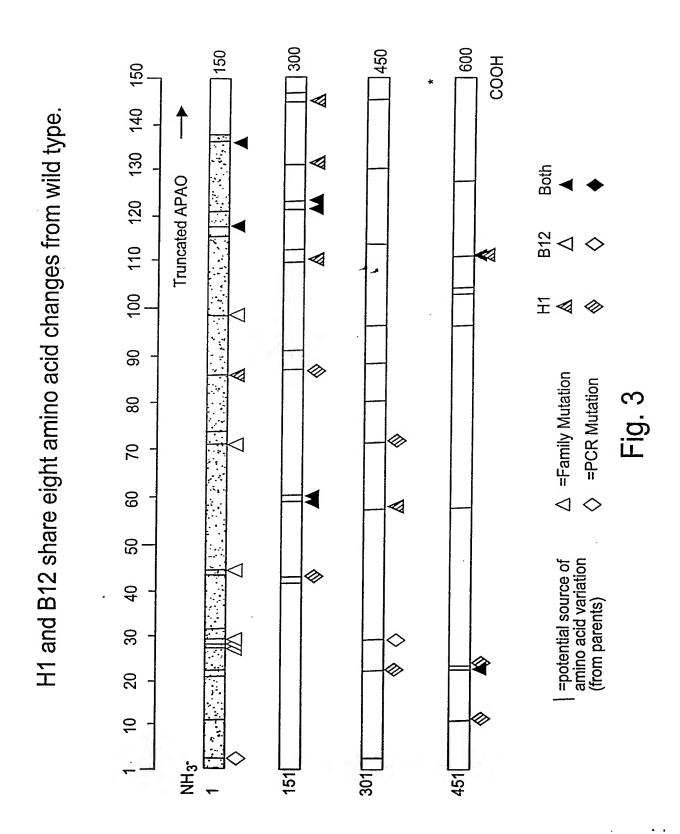
H1 and B12 compared to wild type APAO

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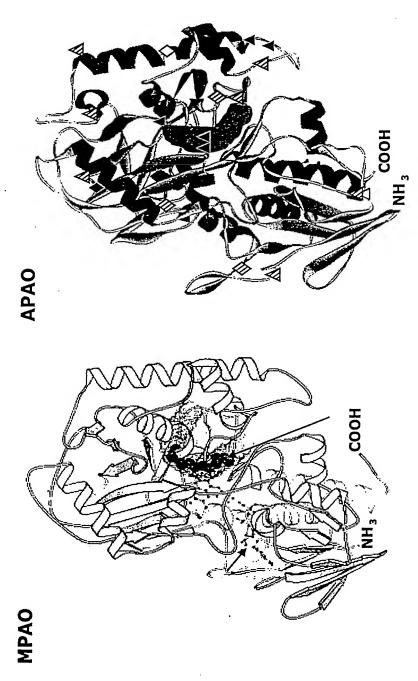
Fold Impr kcat /km 0.0 AP1, pH 5.5 430.0 544.0 280.0 km 662.0240.0 200.0 kcat 7.4 26.1 Fold Impr 40.0 7.3 kcat /km FB1, pH 5.5 70.0 98.0 62.0 Km 2800.0 150.0 701.0 kcat Variant B12

Fig. 2

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Several H1 & B12 mutations map to a putative substrate binding region of APAO



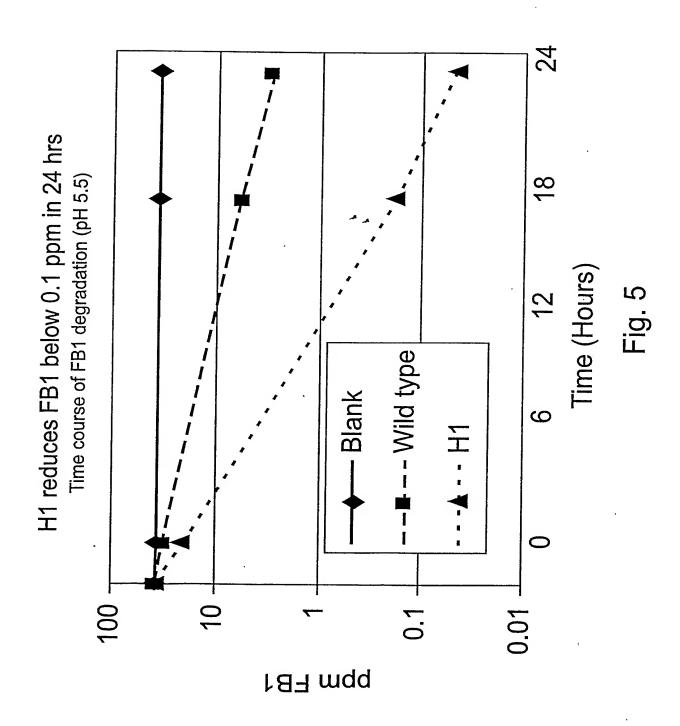
Maize Polyamine Oxidase (MPAO) 30 A crystal structure (Binda et al. 1999. Structure 7:265) Substrate "tunnel" shown in wire form.

APAO (truncated, amino acids 142 -600) 3-D Model after Binda et al. Putative substrate "tunnel" shown in center right.

Mutations: B12  $\triangle \lozenge$  H1  $\triangle \lozenge$  Both  $\blacktriangle$ 

Fig. 4

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H1 retains its high substrate specificity for fumonisins

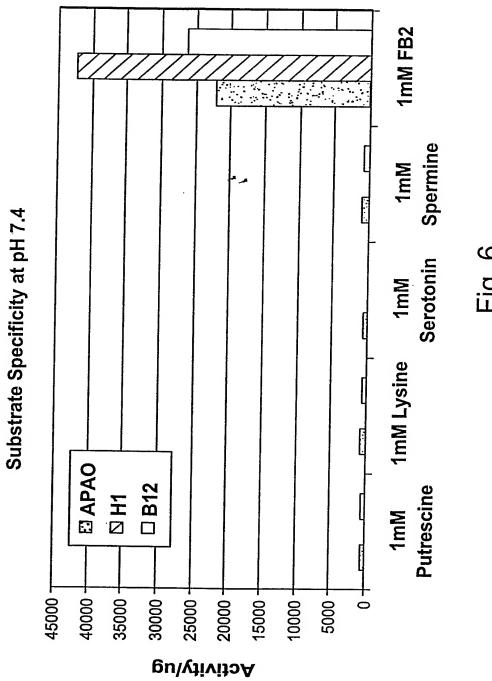
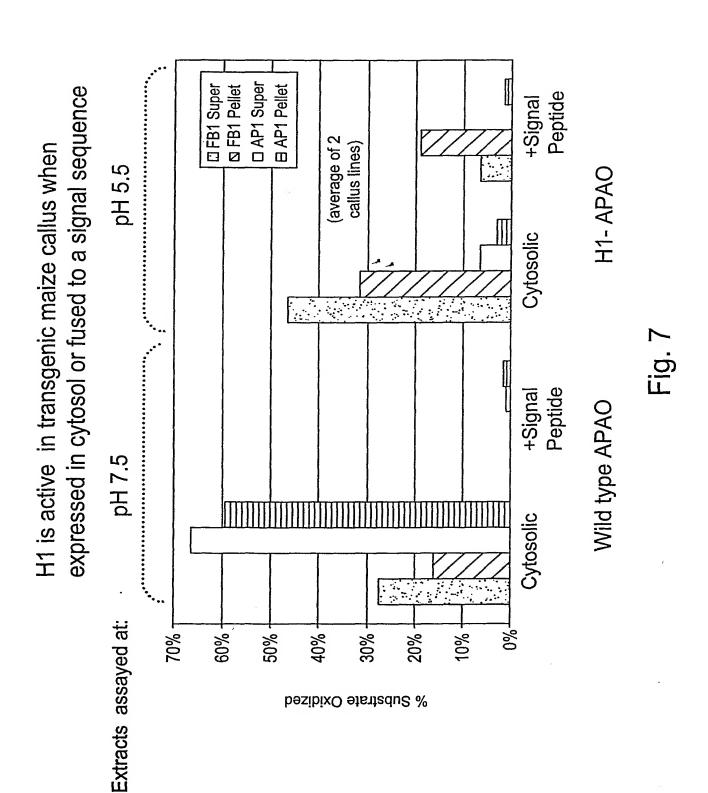


Fig. 6



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Fig. 8A

	riwmitvug (prior to preincubatio	n) %activity (after preincubat
· h1	1325	0.21
<u>q6</u>	1804	0.59
1b6	1612	0.55
1h8	1804	0.35
3e7	1558	0.71

Fig. 8B

	km	kcat
h1	259.12	2164.50
g6	154.19	1612.90

Fig. 8C

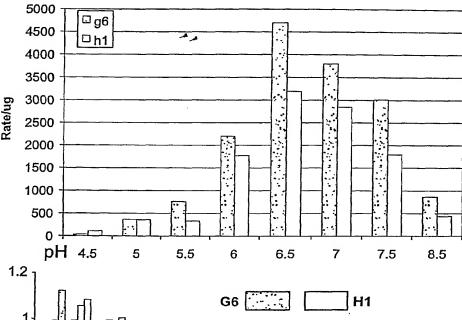
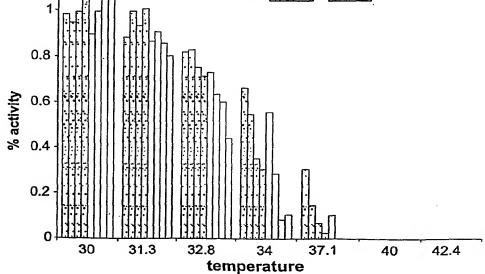
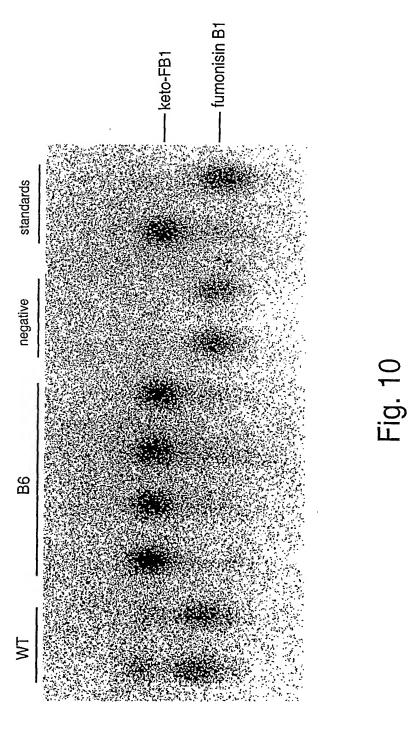


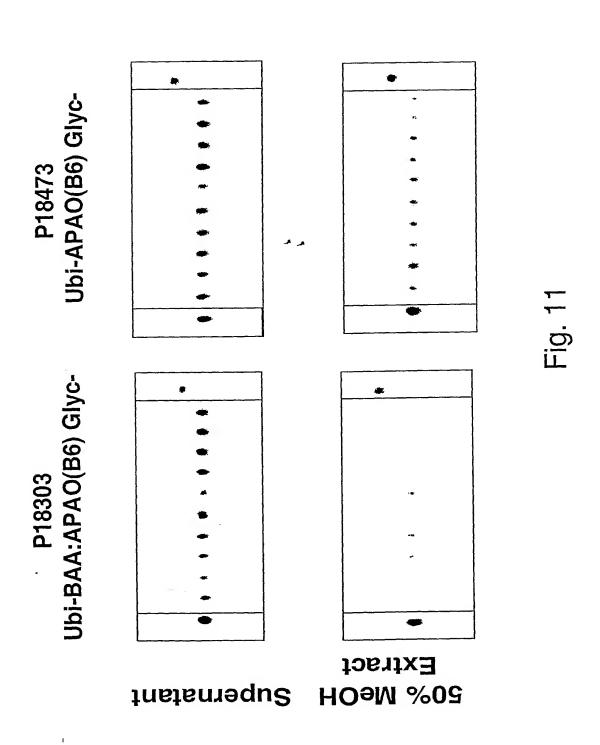
Fig. 8D



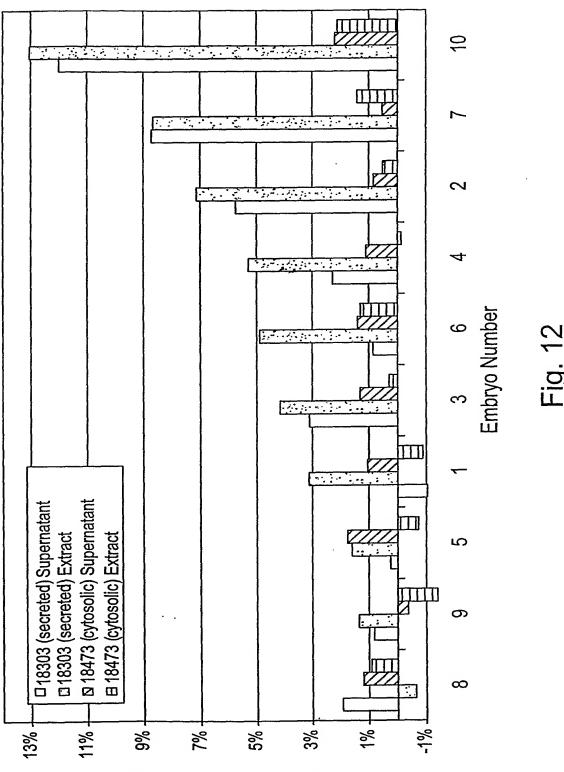
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% Conversion (minus bckgd)

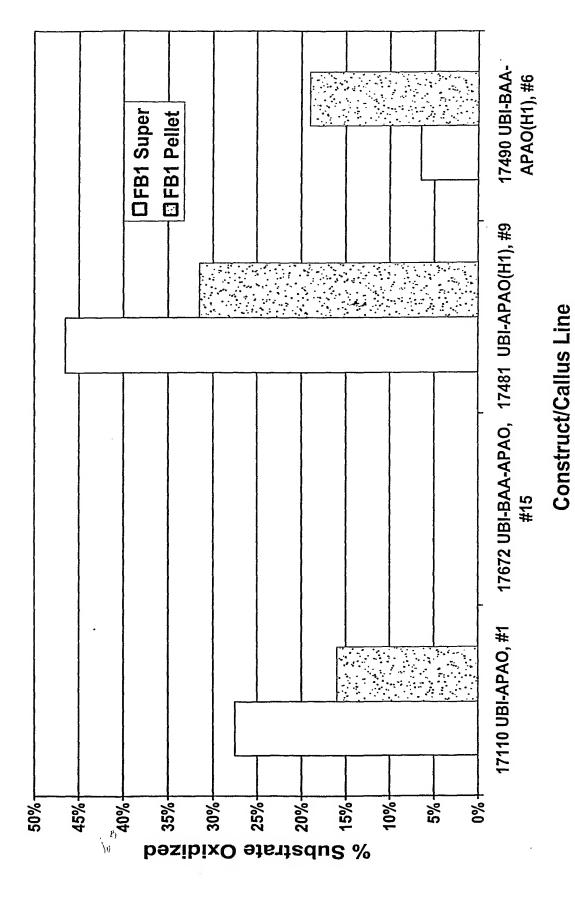


Fig. 13